

MICROBIOLOGY LABORATORY (BIOL 310L) SCHEDULE

Spring 2017

Lecture Professor: Dr. Susan Morrison
Lab Instructors: Ms. Tracy Hirsch
Dr. Susan Morrison

Required:

- (1) Leboffe & Pierce, Microbiology Laboratory & Theory Application, Brief, 3rd edition.
- (2) Coursepack for BIOL310 and BIOL310L
- (3) Sharpie marker, Safety Glasses, Lab Coat; Ms. Hirsch also requires a **bound** composition notebook.

“Pack” pages refer to the laboratory portion of BIOL 310 coursepack from SAS-E-Ink. The Leboffe & Pierce manual pages are listed as “Leboffe.” Everything that is listed must be read before coming to class.

I January 23-January 26 INTRODUCTION; SAFETY; ASEPTIC TECHNIQUE; USE OF MICROSCOPES; OBSERVATION OF PREPARED SLIDES; ENVIRONMENTAL SAMPLE; EPIDEMIC

<u>Leboffe</u> pages 1--8; <u>Pack</u> pages L7-24	Safety & Laboratory Guidelines (read thoroughly, understand and apply throughout the semester)
<u>Leboffe</u> Ex. 1-4	Common Aseptic Transfers & Inoculation Methods
<u>Leboffe</u> pages 59-60	Microbial Growth, Ubiquity & Diversity (read)
<u>Leboffe</u> Ex 2-1	Ubiquity of Microorganisms
& <u>Pack</u> pages L29-30	Distribution of Microorganisms in the Environment
<u>Leboffe</u> pages 141-142	Microscopy & Staining (read)
<u>Leboffe</u> Ex 3-1	Introduction to the Light Microscope
<u>Pack</u> pages L25-28	Observation of Prepared Slides of Bacteria (3 bacterial morphologies at 3 magnifications each)
<u>Leboffe</u> Ex. 7-4	Epidemic Simulation (time permitting)

Notes:

- *+ indicates exercise for which follow-up will be necessary. The time in brackets [] indicates the approximate time span at which follow-up should be done.*
- *⁹“Pack” pages refer to the BIOL 310 coursepack from SAS-E-Ink. Leboffe refers to the manual by Leboffe & Pierce.*
- *Appendix = an appendix in the lab manual. You should familiarize yourself with it, but do NOT memorize it. It is for reference only.*
- ***NOTE:**In addition to the questions for exercises which you do, you are also responsible (on tests and in your notebooks) for questions in exercises requiring only reading and for results and questions for exercises done as demonstration. You are also responsible for all parts of the exercises done from the coursepack or handouts, as well as from the lab book.*
- *Important Note: If the schedule needs to be shifted because of class cancellation for a hurricane, influenza, or other emergency during the term, the date of the lab final and/or other activities may change.*
- *Lab books may be collected and graded at ANY time during the semester; this could occur once or more than once and may be announced OR unannounced. You should come to class at all times with your lab book(s) organized, complete and up-to-date.*

II January 30-February 2 ASEPTIC TECHNIQUE, ENVIRONMENTAL SAMPLE & CULTURE CHARACTERISTICS (continued); PREPARATION OF SLIDES & OBSERVATION OF SIMPLE STAINS & NEGATIVE STAINS; STREAK PLATES; PREPARATION OF CULTURE MEDIA; THE AUTOCLAVE

<u>Leboffe</u> Ex. 1-5 & <u>Pack</u> L36-37	Streak Plate Methods of Isolation
<u>Leboffe</u> Ex. 1-4	Review Common Aseptic Transfers & Inoculation Methods

<u>Leboffe</u> pages 173-176	Bacterial Structure & Simple Stains (read)
<u>Leboffe</u> Ex 3-4	Simple Stains (includes making a bacterial smear)
<u>Leboffe</u> Ex 3-5	Negative Stains

Listing for this day continued on next page

II January 30-February 2 (continued)

<u>Leboffe</u> page 87 (Top)	Environmental Factors Affecting Microbial Growth
<u>Leboffe</u> Ex 2-1	Continue: Ubiquity of Microorganisms
& <u>Pack</u> L29-30	Continue: Distribution of Microorganisms in the Environment
<u>Leboffe</u> Ex 2-2	Colony Morphology
<u>Leboffe</u> Ex. 2-3	Growth Patterns on Slants
<u>Leboffe</u> Ex. 2-4	Growth Patterns in Broth
<u>Leboffe</u> Ex 1-3	Nutrient Broth & Nutrient Agar Preparation [read & understand; we will not be able to carry this out because our temporary lab is not adequately equipped.]
& <u>Pack</u> L31-33	
<u>Leboffe</u> Ex 2-5	Evaluation of Media (read only)
<u>Leboffe</u> Ex 2-11	Steam Sterilization (read only)

III February 6-9 PATHOGEN POSTER PROJECT SIGN-UP; BEGIN IDENTIFICATION OF "UNKNOWN" BACTERIAL CULTURE; GRAM STAIN; USE OF SPECIAL PURPOSE MEDIA (SELECTIVE & DIFFERENTIAL); ISOLATION OF PURE CULTURES

<u>Pack</u> L57-62	Sign up & begin pathogen poster project ---Organize teams of 4 students; select pathogen & area of body for normal microbiota
<u>Leboffe</u> Ex. 1-5 & <u>Pack</u> pp. L36-37	Streak Plate Methods of Isolation (use for Unknown culture)
<u>Leboffe</u> page 545	Identification of Unknowns (read)
<u>Pack</u> L38-L50	Identification of Unknown Bacterial Cultures
<u>Leboffe</u> Ex 3-12	Morphological Unknown (read)
<u>Leboffe</u> page 187 (top)	Differential & Structural Stains (read)
<u>Leboffe</u> Ex. 3-6	Gram Stain
<u>Leboffe</u> pp. 227-229	Selective Media & Differential Media
<u>Leboffe</u> Ex. 4-1	β-phenylethylalcohol Agar
<u>Leboffe</u> Ex. 4-2	Columbia CNA Agar (read only, pay particular attention to figure)
<u>Leboffe</u> Ex. 4-3	Mannitol Salts Agar
<u>Leboffe</u> Ex. 4-4	MacConkey Agar
<u>Pack</u> L60	CLED Agar
<u>Leboffe</u> Ex 2-2	Cultural Characteristics of Microorganisms & Colony Morphology Use this to describe Environmental Samples & for all future observations
<u>Leboffe</u> Ex 2-3	Growth Patterns On Slants
<u>Leboffe</u> Ex 2-4	Growth Patterns In Broth
<u>Pack</u> L34-35, L44	Dichotomous key practice —begin today (& continue in subsequent weeks)

IV February 13-16 DETERMINATION OF OXYGEN REQUIREMENT; METHODS FOR GROWING ANAEROBES; BIOCHEMICAL TESTS; HYDROLYTIC ENZYMES; CATALASE; NITRATE TEST; SPORE STAIN

DEADLINE: for **pure working cultures** of your unknown; **reserve & working** culture slants of unknown

<u>Leboffe</u> page 95 (top)	Aerotolerance (read) & Oxygen Requirements
<u>Pack</u> pages _____	Determination of Atmospheric Oxygen Requirements (using Agar Deep Stabs)
<u>Leboffe</u> Ex 2-6	Fluid Thioglycollate Medium for culturing anaerobes [Read]
<u>Leboffe</u> Ex 2-7	Anaerobic Jar for culturing anaerobes [Read]

<u>Leboffe</u> pages 267-270	Differential Tests (Read)
<u>Leboffe</u> Ex. 5-4	Catalase Test
<u>Leboffe</u> page 331	Tests Detecting Hydrolytic Enzymes (read)
<u>Leboffe</u> Ex. 5-10	Starch Hydrolysis (Amylase Test)
<u>Leboffe</u> Ex. 5-13	Casein Hydrolysis (Caseinase or Casease Test)
<u>Leboffe</u> Ex. 5-6	Nitrate Reduction Test [24-48 hr] (Biochemical Test)

Continue or complete ongoing exercises & identification of unknown cultures

<u>Pack</u> L34-35, 44	Continue Dichotomous Key practice
<u>Pack</u> L57-62	Continue group work on Pathogen Poster & Normal Microbiota

V February 20-23 BIOCHEMICAL TESTS—SUGAR FERMENTATIONS; OTHER BIOCHEMICAL TESTS (IMViC); SULFIDE; MOTILITY (using a SEMI-SOLID AGAR); CONTROL OF MICROBIAL GROWTH---with ULTRAVIOLET LIGHT

<u>Leboffe</u> pages 267-270	Differential Tests (Review)
<u>Leboffe</u> page 279 (Top)	Fermentation Tests
<u>Leboffe</u> Ex. 5-2	Carbohydrate Fermentation using Phenol Red Fermentation Broth [24hr]
<u>Leboffe</u> Ex. 5-19	Carbohydrate Fermentation using Triple Sugar Iron Agar **[18-24hr]**
	IMViC Test Battery
<u>Leboffe</u> Ex. 5-3	Methyl Red Test
<u>Leboffe</u> Ex. 5-7	Citrate Test
<u>Leboffe</u> Ex. 5-18	SIM Medium: Motility, Indole and Hydrogen Sulfide Test
<u>Leboffe</u> Ex. 2-12	The Lethal Effect of Ultraviolet Radiation on Microbes

<u>Pack</u> L34-35, 44	Continue Dichotomous Key practice
	Continue or complete ongoing exercises & identification of unknown cultures
<u>Pack</u> L57-62	Pathogen Poster/Normal Microbiota Project continued

IMPORTANT SCHEDULE NOTE: You may need to return to the lab the next day (ideal) or the day after to read these test results. If reading of the results is delayed and they can't be properly stored, they won't be accurate. Your lab instructor will give you directions.

VI February 27-March 2 ACID-FAST STAIN; ENDOSPORE STAIN; BIOCHEMICAL TESTS; MOTILITY (using WET MOUNTS); BIOCHEMICAL I.D. SYSTEMS

<u>Leboffe</u> Ex. 3-7	Acid Fast Stain (a differential stain)
<u>Leboffe</u> Ex. 3-9	Endospore Stain (a structural stain)
<u>Leboffe</u> Ex. 3-10	Wet Mount & Hanging Drop Preparations
<u>Leboffe</u> Ex. 9-5	Identification of Enteric Microorganisms Using Computer-Assisted Multi-Test Microsystems (demonstration)
<u>Leboffe</u> L71-74, <u>Pack</u> L57-L62	Pathogen Poster project continued
<u>Pack</u> L34-34, 44	Continue Dichotomous Key practice
<u>Pack</u> L38-50	Continue or complete ongoing exercises & identification of unknown cultures
-----	Continue identification of unknowns [See message below about media requests.]
	Review for next week's test

*****Friday March 3--noon; Monday, October 10-noon--Deadline** for requesting supplemental media for unknown culture identification.*** You may request new media not previously used, and will be advised whether it can be provided. All requests must be in writing or by e-mail to your instructor using the subject line : Special Media Request. Please explain why this medium is of value for identification of your unknown. For previously used media, you should indicate why it is necessary for you to repeat the test now if you did not repeat a test immediately after first reading the results. It may take 3-4 days to get these media prepared. Forgetting to come in to read test results is NOT a valid reason to request more media. **HINT:** Request media sooner than this date to permit more time to apply those results.

VII March 6-9 Spring Break-Lab Will Not Meet

VIII March 13-16

***** ****LABORATORY TEST (closed book)**** [practical set-ups & written only sections]
 ***** ****PRACTICAL TESTS IN ASEPTIC TECHNIQUE; PLATE STREAKING; MICROSCOPE FOCUSING****
 ----- Continue or complete ongoing exercises & identification of unknown cultures

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IX March 20-23 Work on Pathogen Poster Project; Dilutions; SENIORS HAVE SPECIAL ASSIGNMENT

Special Assignment for Seniors

<u>Posters</u> L57-L62	Continue work on pathogen poster & normal microbiota project with your team
<u>Leboffe</u> Ex. 7-3	Morbidity & Mortality Weekly Report (MMWR) Assignment (use your pathogen poster microbe)
<u>Pack</u> pp. L63-72	Dilutions Tutorial [to prepare for Viable Count (Standard Plate Count) next week]

